REVIEW ARTICLE

SEMIOCHEMISTRY IN HIGHER ANIMALS: PERCEPTION AND EXPRESSIVE PHYSIOLOGY

Mousumi Das

Department of Botany, Vivekananda College for Women, Kolkata-700008 Correspondence: dassumi222@gmail.com

Abstract: Odor biology has accomplished conformist role in shaping co-evolution of plantanimate existence of the planet ensconced upon inter communicating linkages other than audiovisual reciprocation. Semiochemicals comprising significant fractionation of Volatile Organic Compounds (VOCs) constitute pool of natural odor be classified as pheromones and allelochemicals with subclasses. Objective of the review lies in pointing out brief insight of the formative and transducing mechanism for conventional plant originated volatile molecules (not aiding the purpose of sex) in higher animals and manifestation of inhalation in their physiological kinesics emphasizing onto cat (Felis catus) attractant molecules from various plant families discovered till present along with comparative behavioral analysis. The illustrative methodology of olfaction in vertebrates and higher brain structure for semiochemicals of plant origin frame the distributary pathways of odorant perception via main olfactory bulb for conventional VOCs and accessory olfactory organs like vomeronasal organ (VNO) for pheromones entrusting G-protein coupled receptor (GPCR) dependency especially in mammals. The numbers of intron-less coding sequences for olfactory receptors (OR I and OR II) considerably reduces in high end mammalian evolutionary stem leading to pseudogene constitution. The decade old story of distinct behavioral concoction found in subfamilies of Felidae with response to cis- trans configured active compound (Nepetalactone) from the genus Nepeta and matatabilactone, actinidine like hallucinogen from species other than Nepeta was correlated with recent discovery of prolonged drug like response of cat to iridoid compounds (Isodihydronepetalactone and isoiridomyrmecin) and seven Gas-Chromatography (GC) identified Non-steroidal anti- inflammatory drug (NSAID) compounds from non-aerial portion of Acalypha indica plant. With the analysis of unit behavioral aspect elaborated in literature and performed bioassay, it was found that though the reactive function to the latter was analogous to 'catnip response', the exact pathway for olfactory signal transduction yet not clear especially for character like the genital licking which was not found earlier and also the same to be tested for other members of Felidae as well to draw the continuum.

Key words: Semiochemicals, conventional VOCs, olfaction, GPCR, catnip response, Acalypha indica, NSAID

Communicated:18.06.2024

Revised:30.08.2024

Accepted:15.09.2024

1. INTRODUCTION

Following chemotactic principle of motility for survival in living kingdom, Semiochemicals have established a pool of informatory molecules carrying high end potentiality for intraspecific and interspecific communications panning wide range of species in the ecosystem. The mechanism of signaling by producing chemical attractant or deterrent establishes a third channel for plant-plant, plantanimal or animal-animal interaction beyond audiovisual trajectory of responsiveness. Semiochemicals could be classified in broader groups of Pheromones (intraspecific) and allelochemicals (interspecific) where in the latter be sub-grouped further into Allomone, emitter friendly and Kairomone, receiver friendly in nature [1]. Many-a-times, these signaling molecules are volatile in nature with lowmolecular weight and comprises considerable percentage of Volatile Organic Compounds (VOCs). The science of odor has shaped wide array of communication taking from host-parasite interaction, induction of behavioral changes in pollinator insects and aphids, direct and indirect defense mechanism in plants, pest controlling agriculture [2] to mother-child interrelation, acceleration of puberty and block pregnancy, mediating as mighty sex attractant in higher animals [1], which contributed in structuring plant-animal co-evolution since time immemorial framing the ecological skeleton. Though the scientific repository provides illustrative studies on insect behavior in response to chemical signals from plants, such studies are less in comparison in case of higher animals or mammals in particular. Deterrent effect of monoterpenes like camphor and α -pinene on *Flourensia cernua* DC (tarbush) pellet consumption by sheep explained differential herbivory impacted by individual volatile compound or amalgamated form produced by shrubs [3, 4]. The study comprised of spraying selected chemicals such as α -pinene, camphor, limonene, cis-jasmone, β -caryophyllene and borneol individually on Alfalfa pellets for sheep consumption. Long back, induction of P450 oxidases in rat was established with response to a-pinene and borneol [5]. Probably such exclusive detoxification mechanism retained monoterpene consumption "Generally Recognized as Safe" for the mammals while toxic to insect community [6]. However, Estell's study revealed production of few leaf monoterpenes led to differential use of Alfalfa in selection of diet by the ruminants. Deviated adaptive mutualistic behavior of few Nepenthes sp. from simple carnivorous food habit have been shown just to accommodate the need of specific vertebrate species revealing unpredicted avenue for plant-animal coevolution [7]. Nepenthes hemsleyana, found in low land forest of Brunei Darussalam has developed alternate pitcher with elongated tube and low-level fluid to provide roosting space for *Kerivoula hardwickii* (woolly bats) [8, 9]. Similarly, N. lowii produces white jelly-like secretion that attracts Tupaia montana (tree shrews) [10]. Also, it has been reported that N. rajah, another Nepenthes species of higher mountain has been adopted itself to successfully capture faeces of T. montana and Ratus baluensis (summit rats) in its pitcher by virtue of their positioning while accessing the nectar [11,12]. The story of cat attracting plant Nepeta cataria is well known to humanity for decades. Such behavioral attribute called "catnip response" has been proved to be a manifestation of Nepetalactone, an insect repellent as the first identified cat attractant chemical from the plant reported but without performing bioassays [13]. Later an unpublished research work [14] reported a group of lactones containing epinepetalactone, isodihydronepetalactone, iridomyrmecin, neonepetalactone. dihvdronepetalactone and isoiridomyrmecin along with nepetalactone itself eliciting such response in domestic cats. This interesting plant-mammal interaction indulged many researchers in recent time to investigate if any other cat attractant plant present in nature.



Figure 1. tree shrew (*Tupaia minor*) collecting nectar from the lower lid surface of *Nepenthes gracilis* pitchers [7] with a gesture of positioning their faeces inside the altered designed pitcher.

Mechanism of olfactory transduction for general odorants

Science of olfaction constitute an obvious pathway of plant-animal coevolution by means of odor perception mediated by transducers and amplifying receptors. Plant odorants are integral component of semiochemistry though their mechanism of functionality differs in the range of animal taxa. Plant-pollinator relationship was well established conventionally as a derivative of odor biology [15] where it is indispensable to manifest important life cycle phenomena sometimes involving volatile semiochemicals of floral microbiome too, aiding the methodology of perception for insect community [16] but the nitty gritty for complex processing of any plant originated odorant in higher brain structure of the vertebrates and its expressive concoction brings the nuance in the study. In mammals, such perception of plant volatile ligand is occurred by Odorant Receptors (ORs), the largest family of heptahelical [17, 18] G-protein coupled receptor that spans the epithelial layer of Olfactory Sensory neurons (OSNs) in the posterior nasal cavity and is responsible for strong olfactory activity in mammals by converting chemical information into electric impulse and opening of nonselective cation channels with production of cyclic AMP (cAMP) pool [19] and neuron depolarization [20]. OSNs are bipolar neurons with dendritic end in nasal cavity and axon in olfactory bulb of higher brain [21, 22]. Odorant Binding Proteins (OBPs) containing lipocalins with β - barrel foldings [23] play significant role in passive transportation of hydrophobic odorant ligands through epithelial mucus layer by formation of OBP-odorant (ligand) complex [24] that selectively detected by olfactory neurons [25-27]. Whereas the epithelial activity of OSNs is an established hypothesis of first line response for smell sensitization in higher animals especially in mammals [28, 29], many advanced studies have demonstrated the chemosensory process to be more complex and tightly regulated [30, 31]. Trace amine-associated receptors (TAAR) expressed on olfactory epithelium are able to sensitize odorants containing volatile amines [32], similarly some members of guanylyl cyclase D(GC-D) expressing OSNs [33, 34] of dorsal olfactory bulb (OB), sensory neurons in the septal organ (SO) of nasoplatine duct [35, 36], mature Grueneberg ganglion (GG) of olfactory bulb (OB) expressing olfactory marker protein are functional instruments [37] for odor discrimination from the natural pool.

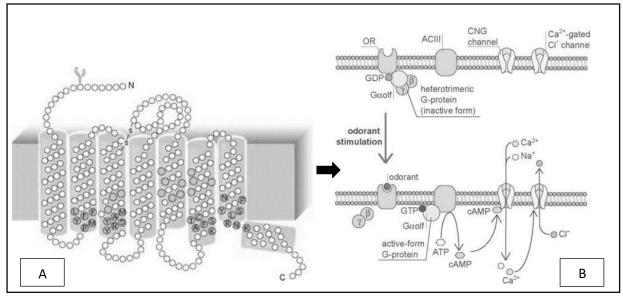


Figure 2. Canonical pathway of olfactory signal transduction in OSNs[B] via activation of heptahelical Olfactory Receptors [B] spanning posterior nasal cavity of mammals [22].

Table1. Cell signaling pathways responsible for transduction of some common plant originated				
volatile molecules [48]				

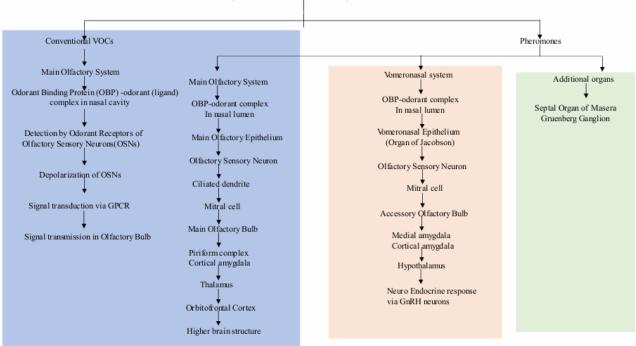
Sl No.	Plant Volatiles (VOCs)	Source Plant	Regulator Pathway After Detection
1	Zerumbone	Curcuma zerumbet	Downregulating hedgehog (Hh/GLI) signaling pathway
			• Upregulating TNF-related apoptosis-inducing ligand pathway
			• Interference in NF-κB signaling Pathway
2.	Phytol	Common plants	• Interferon (TNF-β) signaling pathway
3.	β-caryophyllene	Edible plants	• Selectively binds with cannabinoid type-2(CB ₂) receptor
			• Inhibition of toll-like receptor complex CD14/TLR4/MD2 pathway
4.	Carvacrol	Unknown	 Activation of peroxisome proliferator-activated receptor gamma (PPAR-γ)
			• Downregulation of COX-2 pathway
5.	Thymoquinone	Nigella sativa L.	Active in cyclooxygenase and
			lipoxygenase activity
6.	D-Limonene	Citrus fruit	• Binds with transient receptor potential melastatin- 8(TRPM8) described as prostate cancer marker

Mechanism of olfaction had been well studied for decades in insect model organism *Drosophila melanogaster* [38], worm *Caenorhabditis elegans* [39] and zebrafish *Daniorerio* [40] amongst which insect model attracts highest interest where antennal lobe behaves as equivalent of the Olfactory Bulb to the mammals. Here, both antenna and maxillary pulp [41, 42] containing the sensory hairs called sensilla [43, 44] express Olfactory Receptor Neuron (ORNs) responsible for transduction of chemical information to higher brain structure like protocerebrum [45], therefore, constituting a G-protein coupled receptor (GPCR) independent pathway of scent perception [46]. Despite of bearing absolute different architecture for olfactory receptor organs, higher mammals (human) and insects show commonness in detecting volatile enantiomers of carvone, menthol, D-limonene, α -pinene etc. Similarly, S-(+)-carvone is sensed like caraway whereas R-(-)-carvone is detected like spearmint in human [47]. However, perception of odorants other than VOCs again constitute an GPCR pathway where Vomeronasal Organ (VO) acts as mediator to carry forward chemosensory response from Accessory Olfactory Bulb (AOB) to higher brain structure [21, 22].

2. GENETICS OF ODORANT RECEPTORS

Discovered in rat for the first time with extreme diverse amino acid sequences [49], Olfactory Receptor (OR) genes are also widely dispersed in mammal genome majorly into tandem cluster [50] and are intron less. They are encoded by multigene family ensuring higher diversity and broad distribution [51] except chromosome 18 and Y in mouse [52, 53] and chromosome 20 and X [54]. Depending on sequence and evolutionary track, the OR genes are categorized into two separate classes like OR I and OR II genes. 80-90% OR genes in mammals is made up of class II [55]. Though OR I genes were thought to be able to discriminate water-soluble odorants only with exclusive distribution in catfish [56] and amphibians [57], their presence was reported later in mammals too [58]. These genes contain ~1kb coding region with OR sequence motifs [59]. Number of intact OR genes vary considerably in different mammal species from ~1000 in mouse and ~800 in dog to ~370 in human [59] and those who are unable to code functional OR proteins are termed as OR pseudogenes. However, the olfactory sensitivity in higher animals is much correlated with number of glomeruli present on each Olfactory Bulb rather than total number of intact OR genes in organism [60] and quantity of functional OR gene decreases in higher evolutionary stem [61].

The receptive mechanism of semiochemicals like pheromone by higher group of animals is initiated primarily by G-protein coupled receptors of V1R (Vanilloid type 1 receptor) and V2R (Vanilloid type 2 receptor) on vomeronasal epithelium. While V1Rs expressing $G_{\alpha 12}$ are dominant in the apical neurons of vomeronasal organ, V2Rs are explicitly located in the basal neurons of the same with expression of G α o subunit. Presence of functional V1R receptors have significantly diminished in higher group of mammal e.g., human genome consists of several V1R pseudogenes except only five functional genes. Dogs are able to perform exceptionally well functioned olfaction with only eight functional V1R genes. Phylogenetic analysis shows both of the gene families emerged to complement the basic olfactory requirements in rodents, however they underwent countable gene losses in carnivores in long run and also multiple mutations in their subfamilies to the level of non-functional pseudogenes [62].



Perception of odorants in higher animals

Figure 3. Schematic presentation of olfaction in higher animals [62]. Color blocks represent division of olfaction like Main Olfactory Bulb is involved in transmission of both conventional VOCs and pheromone (blue colored); Vomeronasal Organ (VNO), Septal Organ of Masera (SO) and Gruenberg Ganglion (GG) are involved in olfaction of pheromone (red and green block).

3. BIOSYNTHETIC PATHWAYS FOR CONVENTIONAL PLANT VOLATILES

Unlike plant produced primary metabolites used in growth, development and reproduction, the volatile compounds including semiochemicals comprise major percentage (~1%) of plant secondary metabolites [63] that play significant role in ecological communication. In addition to plants, major metabolic pathways in human, microbes (mVOC) and other animals are also responsible to emit airborne volatile compounds [2]. VOCs are sequestered carbons of lower molecular weight with higher vapor pressure for which cellular membranes of living organism show high permeability [64]. Being lipophilic in nature [63], the resultants of Lipoxygenase (LOX) pathway [48] include release of hydrophilic counterparts, reduction, hydroxylation/oxidation, methylation, acetylation reaction to furnish their biosynthesis [64] and are classified in distinct groups like terpenoids, alkaloids, carotenoids, fatty acid derivatives, phenyl propanoids and several amino acid derivatives. Four pathways namely mevalonate pathway (MVA), shikimate pathway, Methyl erythritol (MEP) and acetate pathway with spatial subcellular localization are responsible for origin of pool of volatile compounds. Amongst them only MEP pathway explicitly marks its presence in plastid with all its enzyme subset [65] that give rise to mainly monoterpenes, hemiterpenes and diterpenes. The MVA pathway considered for development of triterpenes and sesquiterpenes is conventionally localized in cytosol whereas some present study reports its distribution within cytoplasm,

peroxisome and endoplasmic reticulum [66, 67]. Both these pathways contribute isopentyl diphosphate (IPP) and dimethylallyl diphosphate (DMAPP) [68], the two five carbon isoprene building blocks those units for synthesis of further precursor molecules in the pathway like geranyl diphosphate (GPP) for monoterpenes, farnesyl diphosphate (FPP) for sesquiterpenes and geranylgeranyl diphosphate (GGPP)for diterpenes [69]. The precursors of shikimate pathway originate from pentose phosphate and glycolytic pathway to the formation of Phenylalanine (Phe) for giving rise to phenylpropanoid and benzenoid group of VOCs [70]. Though Phenylalanine formation occurs inside plastid [71], rest of the pathway is localized in cytosol. The group of fatty acid derivative volatile compounds are generated from Acetyl-CoA, the end product from glycolytic pathway through lipoxygenase (LOX) biochemical reactions using 13-Hydroperoxylelonic acid as an intermediate compound [72].

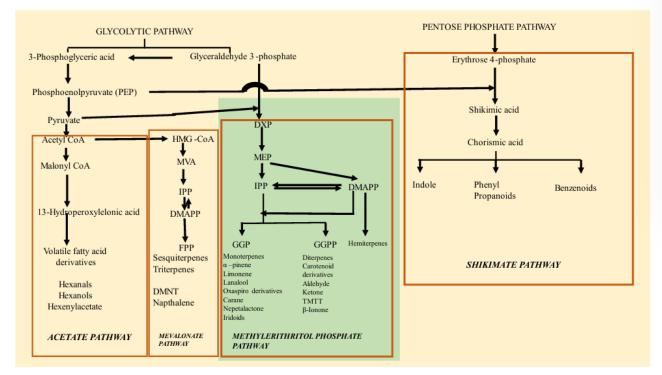


Figure 4. Schematic representation of biosynthetic pathway of plant Volatile Organic Compounds (in brief) with spatial organization in cellular compartments [2, 48, 63] mentioning formative pathways for cat [86, 90] and sheep attractant molecules [3]. Color blocks represent cellular compartments of plastid (green block) and cytoplasm (yellow block)



Figure 5. Hallucinogenic response to non-aerial portion (root) of *Acalypha indica* found in feral cats (*Felis catus*) similar to catnip [86].

4. PLANT-HIGHER ANIMAL COMMUNICATION FOLLOWING THE TERMS OF PLANT ODORANTS

Researches have revealed plant originated volatile compounds establishes direct liaison with stimulated behavioral responses in higher animals. The fundamental and pioneering credit of findings in the direction must go to the discovery of Nepeta cataria (catnip) elicited response in Felis catus (domestic cat) almost 200 years ago [73]. The term 'catnip responses' include a group of behavioral expression [Table 2] independent of any sexual aspect [74] regulated by autosomal gene. Such kind of response had also been tested in wide range of animals within subfamilies of Felidae for which positive results were found in Pantherinae subfamily only [75] in Panthera leo (lion), Panthera tigris (tiger), Panthera pardus(leopard), Panthera onca (Jaguar), Panthera uncia (snow leopard) and in Neofelis nebulosa (clouded leopard) amongst which lions and jaguars were found to be extremely sensitive to catnip [76]. Chemical study of the plant revealed more than one molecule with cat attracting property, the principal compound being nepetalactone (cis-trans isomer, 70-99.9%) and epinepetalactone (trans-cis isomer, 0.1-30%) of essential oil obtained from catnip [77]. Differential combination of these two molecules was found in other species of Nepeta like Nepeta nepetella L., Nepeta sibthorpii Benth. and N. hindostana Haines and also in species other than Nepeta like Actinidia polygama, A. kolomikta, Valeriana officinalis L., Teucrium marum L. Menvanthes trifoliata L. and Menvanthes trifoliata L. Amongst these cat attractant molecules, matatabilactone, actinidine [78] and many indole compounds were classified as hallucinogens, potent inhibitors of acetylcholinesterase. Amazingly smoking of catnip was found to be pleasurable for human being [79] with a hint of its hallucinogenic effect [Table 2]. The operation of functionality explores that smell is the sole determinant of catnip originated cat attractant behavior where vomeronasal organ is not

involved directly in the process [80] but the ventromedial nucleus of hypothalamus was found to be responsible to respond to such olfactory stimulation [81]. Interestingly each unit of 'catnip response' is apparently juxtaposed to sexual or ingestive behavior [82] whereas the removal of amygdala expressed no effect on such response to *Actinidia polygama* [83] suggesting a probable cross-reactive analysis of such response with some sort of naturally originated social odor in the family of Felidae like various glandular secretion and odor of urine and feces [84]. Study in recent decade revealing drug like activity of *Acalypha indica* or Indian nettle root on feral cat introduced a new chapter to the story of plant elicited cat attractant molecules wherein the interesting chemical analysis by the researchers has identified (4R,4aR,7S,7aR)-isodihydronepetalactone and (4S,4aS,7S,7aR)-iridomyrmecin from *Acalypha indica* dried root those matched with prepared synthetic hydrogenated oil from *Nepeta cataria* containing *cis*-fused and *trans*-fused nepetalactone [85]. Also, an Indian patent entitled 'A METHOD FOR ISOLATING THE CHEMICAL COMPOUNDS FROM THE FRESH ROOTS OF *ACALYPHA INDICA* LINN' [86] has identified group of hallucinogenic, anti-spasmatic attractants of cat. Strong olfactory activity is well known in pig for distinguishing social odor, utilizing such recognition for reproduction [87, 88] and also respond to odor from non-social origin has reported [89].

SL No	Molecules	Responses
1.	Odor from Aniseed, Cedarwood, Pine, Thyme, Lavendel, Cinnamon bark, Ginger	Repeated scratching of head and neck, flopping on either side of body with or without rubbing, sniffing in pig. [89]
2.	Alfalfa pellet mixed with volatiles of limonene, jasmone, β-caryophyllene, borneol	Enhancement of consumption by sheep [3].
3.	Smashed leaves of <i>Nepeta cataria</i> (catnip) releasing nepetalactone, epinepetalactone, dihydronepetalactone, isodihydronepetalactone, neonepetalactone, 5,9-dehydronepetalactone	Sniffing the odor, chewing leaves, licking with head shaking, body rubbing, head-over and side by side body rolling, paw licking and salivating, scratching soil [90]. In totality these behavioral expressions are called 'catnip response'.
4.	N. <i>hindostana</i> Haines (native to India) leaves contain traces of nepetalactone	Excites cat [91]
5.	Leaves and galls of <i>Actinidia polygama</i> silver vine) and A. <i>kolomikta</i> containing Matatabilactone which is a mixture of iridomyrmecin and isoiridomyrmecin [92], actinidine and beta-phenylethyl alcohol.	'Matatabi' reaction [82] and salivation in cat

Table 2. Higher animal responses to odor of plant origin

6.	N-(2-p-hydroxy-phenyl) ethyl-	Induce catnip response
	actinidine isolated from dried roots of	
	Valeriana officinalis L. [93];	
	dolicholactone C and D from essential oil	
	of <i>Teucrium marum</i> L. [94];	
	boschniakine and boschnialactone from	
	Boschniakia rossica [95];	
	mitsugashiwalactone from Menyanthes	
	trifoliata L. [95]	
7.	Isodihydronepetalactone and	Induce catnip response
	isoiridomyrmecin from Acalypha indica	
	L. dried root [85]	
8.	Group of seven compounds (1- Oxaspiro	Hallucinogenic, drug-like, anti-spasmatic and anti-
	[2.5] octane, 4,4- Dimethyl 1-8-	inflammatory symptoms in cat apparently similar to
	methylene;1-	catnip response. In addition to this licking of genital
	Napthalenol, decahydro-4a- methyl;	area was recorded by the inventors.
	Carane, 4,5–epoxy-, trans; Naproxen,	
	Nabumetone, Carpofen and Piperidine	
	2,6-dimethyl-1- nitroso) from fresh roots	
	of Acalypha indica L. [86]	
9.	Nepeta cataria smoking through	Makes human happy and helps for intoxication [96].
	cigarettes or pipes or sprayed on tobacco	Has antispasmodic activity in humans.

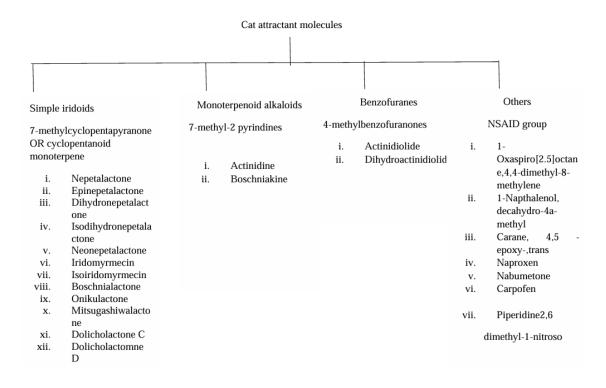


Figure 6. Classification of plant elicited cat attractant molecules [86, 90]

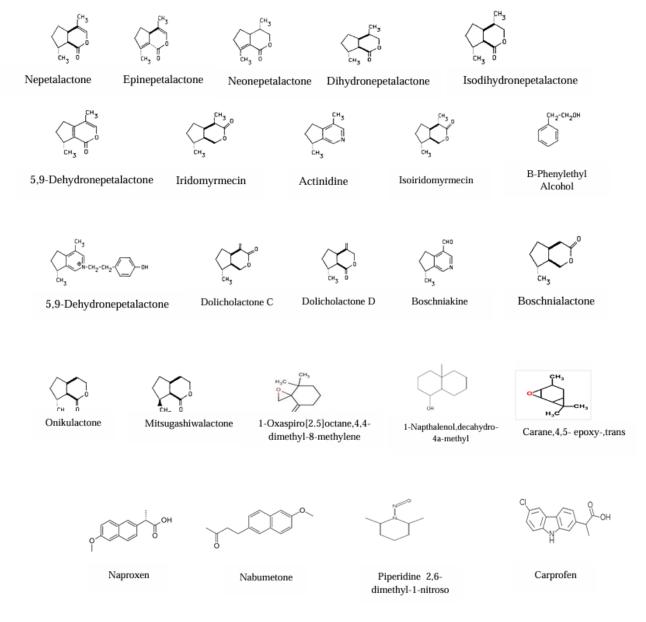


Figure 7. Chemical structure of potent cat attractants discovered till present [85, 86, 90]

5. CONCLUSION

The pool of all natural perfumes (odor) sourced to the environment could be discussed under two broad systems of general odorants and pheromones [97] aiding communication between living organism to its external world. In case of vertebrates and other higher animals, such communication is prerogative of an elaborative mechanism called olfaction, a type of chemo-sensation involving wide array of receptors expressed on peripheral sensory neurons transmitting chemical information to the main olfactory system of brain, principally for general odorants and accessory olfactory system (vomeronasal organ, SO and GG) for pheromone mostly [Figure 2], depending upon the nature of receptors. The odorant molecules function as

ligands to such receptors by forming odorant-receptor complexes could be assessed chemically based on their volatility where the olfactory epithelium is entrusted for detecting highly volatile compounds in relation to the vomeronasal organ [98] that requires direct contact with the source for initiation of olfaction. The unique response to the elicited volatile compounds from catnip by some of the members of Felidae had been demonstrated long back as an activity of main olfactory system, were anesthetized olfactory mucosa [75] and olfactory bulbectomy [80] had abandoned catnip behavior in totality eliminating the notion of vomeronasal organ (VNO) to be connected with such response. As oral administration of active compounds from Nepeta cataria failed to induce 'catnip response' [99], it became thoroughly clear that 'the smell' and not 'the taste' was key to such response to occur [100]. Discovery of similar responses of feral cat to the root (non-aerial part unlike catnip) elicited compounds from Acalypha indica Linn. undoubtedly explores a novel domain in the direction with some differences of identified compounds from dried [85] and fresh root [86] of the plant. The compounds primarily identified from the fresh root determined a strong drug like effect [Table2] as shown in the bioassay which comprises all sort of behavior like catnip except the licking of genital area [86] which was not reported in case of catnip response and also longer interaction time was recorded in comparison to catnip [101]. Now, some unit behavior of catnip response like body rubbing and overhead rolling soon after chewing of the source were conventionally seen as sexual response in cat, thus linking of such behavior to activity of vomeronasal organ was an easy hypothesis to build up until the experiment of olfactory bulbectomy was successfully performed. Therefore, the exact pathway of response against Acalypha root in higher brain structure of cat is yet to establish in order to understand its real nature like whether it is an ingestive response, a pleasure response comprising of sexual unit behavior or mixing of any sort of social odor (saliva, genital secretion, urine etc.) thereto is involved to the process. Presence of Isodihydronepetalactone and isoiridomyrmecin [85] in the dried root system of Acalypha could not merely answer all and full elucidation of this chapter of neuroscience is required to understand the unknown co-evolutionary stems of life.

6. REFERENCES

- Francke, W., "Semiochemicals: mevalogenins in systems of chemical communication", In: Müller, P.M., Lamparsky, D. (eds) Perfumes, Springer, Dordrecht, (1994). https://doi.org/10.1007/978-94-011-3826-0_3
- Dudareva, N., Klempien, A., Muhlemann J. K. and Kaplan, I., "Biosynthesis, function and metabolic engineering of plant volatile organic compounds", The New Phytologist, vol 198, (2013), pp 16 - 32. http://www.jstor.org/stable/newphytologist.198.1.16
- Estell, R. E., Fredrickson, E. L., Tellez, M. R., Havstad, K. M., Shupe, W. L., Anderson, D. M. and Remmenga, M. D., "Effects of volatile compounds on consumption of alfalfa pellets by sheep", Journal of Animal Science, vol 76, (1998), pp 228–233. https://doi.org/10.2527/1998.761228x
- Estell, R. E., Fredrickson, E. L., Anderson, D. M., Havstad K. M. and Remmenga, M. D., "Effect of previous exposure of sheep to monoterpene odors on intake of alfalfa pellets treated with camphor or α-pinene", Small Ruminant Research, vol 58, (2005), pp 33-38. DOI:10.1016/j.smallrumres.2004.08.011
- Oda, Y., Furuichi, K., Tanaka, K., Hiroi, T., Imaoka, S., Asada, A., Fujimori, M. and Funae, Y., "Metabolism of a new local anesthetic, ropivacaine, by human hepatic cytochrome P450", Anesthesiology, vol 82, (1995), pp 214–220. doi: https://doi.org/10.1097/00000542-199501000-00026

- 6. Rice, P. J. and Coats, J. R., "Insecticidal properties of several monoterpenoids to the house fly (Diptera: Muscidae), red flour beetle (Coleoptera: Tenebrionidae), and southern corn rootworm (Coleoptera: Chrysomelidae)", Journal of Economic Entomology, vol 87, (1994), pp 1172–1179. https://doi.org/10.1093/jee/87.5.1172
- Bauer, U., Rembold, K. and Grafe, T. U., "Carnivorous Nepenthes pitcher plants are a rich food source for a diverse vertebrate community", Journal of Natural History, vol 50, (2016), pp 483–495. https://doi.org/10.1080/00222933.2015.1059963
- Grafe, T. U., Schöner, C. R., Gerald, G. K., Anissa, J. and Schöner, M. G., "A novel resource-service mutualism between bats and pitcher plants", Biology Letters, vol 7, (2011), pp 436–439. http://doi.org/10.1098/rsbl.2010.1141
- 9. Schöner, C. R, Schöner, M. G., Kerth, G. and Grafe, "Supply determines demand: influence of partner quality and quantity on the interactions between bats and pitcher plants", Oecologia, vol 173, (2013), pp 191–202. https://doi.org/10.1007/s00442-013-2615-x
- Clarke, C. M., Bauer, U., Lee, C. C., Tuen, A. A., Rembold, K. and Moran, J. A., "Tree shrew lavatories: a novel nitrogen sequestration strategy in a tropical pitcher plant", Biology Letters, vol 5, (2009), pp 632–635. https://doi.org/10.1098/rsbl.2009.0311
- Greenwood, M., Clarke, C., Lee, C. C., Gunsalam, A. and Clarke, R. H., "A unique resource mutualism between the giant bornean pitcher plant, Nepenthes rajah, and members of a small mammal community", PLOS One, vol 7, (2011). https://doi.org/10.1371/annotation/1ee8d7d0-eff5-431f-9059c7dbf1bde347
- Wells, K., Lakim, M. B., Schulz, S., and Ayasse, M., "Pitchers of Nepenthes rajah collect faecal droppings from both diurnal and nocturnal small mammals and emit fruity odour," Journal of Tropical Ecology, vol 27, (2011), pp 347–353. Doi:10.1017/S0266467411000162
- McElvain, S. M., Bright, R. D. and Johnson, P. R., "The constituents of the volatile oil of catnip. I. Nepetalic acid, nepetalactone and related compounds", Journal of the American Chemical Society, vol 63, (1941) pp.1558-1563.
- 14. Nelson, D. L., Methyl cyclopentane monoterpenes of Nepeta cataria and Actinidia polygama, Ph. D. Thesis, Purdue University; 1968.
- Holopainen, J. K. and Blande, J. D., "Molecular plant volatile communication", In: López-Larrea, C. (eds) Sensing in nature, advances in experimental medicine and biology; Springer, New York, NY, (2012), p 739. https://doi.org/10.1007/978-1-4614-1704-0_2
- Crowley-Gall, A., Rering, C. C., Rudolph, A. B., Vannette, R. L. and Beck, J. J., "Volatile microbial semiochemicals and insect perception at flowers", Current Opinion in Insect Science, vol 44, (2021), pp 23-34. https://doi.org/10.1016/j.cois.2020.10.004
- Dixon, R. A., Kobilka, B. K., Strader, D. J., Benovic, J. L., Dohlman, H. G., Frielle, T., Bolanowski, M. A., Bennett, C. D., Rands, E., Diehl, R. E., Mumford, R. A., Slater, E. E., Sigal, I. S., Caron, M. G., Lefkowitz, R. J. and Strader, C. D., "Cloning of the gene and cDNA for mammalian beta-adrenergic receptor and homology with rhodopsin", Nature, vol 321, (1986), pp 75–79. DOI: 10.1038/321075a0
- 18. Jones D. T. and Reed, R. R., "Golf: an olfactory neuron specific-G protein involved in odorant signal transduction", Science, vol 244, (1989), pp 790-795. https://doi.org/10.1126/science.2499043
- 19. Dhallan, R. S., Yau, K. W., Schrader, K. A. and Reed, R. R., "Primary structure and functional expression of a cyclic nucleotide-activated channel from olfactory neurons", Nature, vol 347, (1990), pp 184-187. https://doi.org/10.1038/347184a0
- 20. Firestein, S., Zufall F. and Shepherd, G. M., "Single odor-sensitive channels in olfactory receptor

neurons are also gated by cyclic nucleotides", The Journal of Neuroscience, vol 11 (1991), pp 3565–3572. https://doi.org/10.1523/JNEUROSCI.11-11-03565.1991

- 21. Gaillard, I., Rouquier, S., and Giorgi, D., "Olfactory receptors", Cellular and Molecular Life Sciences CMLS, vol 61, (2004), pp 456-469. https://doi.org/10.1007/s00018-003-3273-7
- Kato A. and Touhara, K., "Mammalian olfactory receptors: pharmacology, G protein coupling and desensitization", Cellular and Molecular Life Sciences, vol 66, (2009), pp 3743-3753. https://doi.org/10.1007/s00018-009-0111-6
- 23. Flower, D. R., "The lipocalin protein family: structure and function", Biochemical Journal, vol 318, (1996), pp 1-14.
- Hajjar, E., Perahia, D., Débat, H., Nespoulous, C. and Robert, C. H., "Odorant binding and conformational dynamics in the odorant-binding protein", The Journal of Biological Chemistry, vol 281, (2006). pp 29929–29937. https://doi.org/10.1074/jbc.M604869200
- Tegoni, M., Pelosi, P., Vincent, F., Spinelli, S., Campanacci, V., Grolli, S., Ramoni, R. and Cambillau, C., "Mammalian odorant binding proteins, "Biochimica et Biophysica Acta (BBA)- Protein Structure and Molecular Enzymology, vol 1482(1-2), (2000), pp 229-240.
- Briand, L., Eloit, C., Nespoulous, C., Bézirard, V., Huet, J. C., Henry, C., Blon, F., Trotier, D. and Pernollet, J. C., "Evidence of an odorant-binding protein in the human olfactory mucus: location, structural characterization, and odorant-binding properties", Biochemistry, vol 41, (2002), pp 7241-52.
- Teixeira, G. D., Esteves, C., Moro, A. J., Lima, J. C., Barbosa, A. J. and Roque, A. C. A., "An odorantbinding protein based electrical sensor to detect volatile organic compounds", Sensors and Actuators B: Chemical, vol 411, (2024), pp 135726.
- 28. Sullivan, S. L., "Mammalian chemosensory receptors", Neuro Report, vol 13, (2002), pp A9-A17.
- Ma, M., "Odor and pheromone sensing via chemoreceptors", In: López-Larrea, C. (eds) Sensing in nature. Advances in experimental medicine and biology; Springer, New York, NY, 2012, 739. https://doi.org/10.1007/978-1-4614-1704-0_6
- Breer, H., Fleischer, J. and Strotmann, J., "Signaling in the chemosensory systems: the sense of smell: multiple olfactory subsystems", Cellular and Molecular Life Sciences CMLS, vol 63, (2006), pp 1465-1475. https://doi.org/10.1007/s00018-006-6108-5
- Munger, S. D., Leinders-Zufall, T. and Zufall, F., "Subsystem organization of the mammalian sense of smell", Annual Review of Physiology, vol 71, (2009), pp 115-140. https://doi.org/10.1146/annurev.physiol.70.113006.100608
- 32. Zucchi, R., Chiellini, G., Scanlan, T. S. and Grandy, D. K., "Trace amine-associated receptors and their ligands", British Journal of Pharmacology, vol 149, (2006), pp 967-78. https://doi.org/10.1038/sj.bjp.0706948
- 33. Liberles S. D. and Buck, L. B., "A second class of chemosensory receptors in the olfactory epithelium", Nature, vol. 442, (2006), pp 645-650.
- 34. Leinders-Zufall, T., Cockerham, R. E., Michalakis, S., Biel, M., Garbers, D. L., Reed, R. R., Zufall, F. and Munger, S. D., "Contribution of the receptor guanylyl cyclase GC-D to chemosensory function in the olfactory epithelium." Proceedings of the National Academy of Sciences, vol 104, (2007), pp 14507-14512. https://doi.org/10.1073/pnas.0704965104
- Kaluza, J. F., Gussing, F., Bohm, S., Breer, H. and Strotmann, J., "Olfactory receptors in the mouse septal organ", Journal of Neuroscience Research, vol 76, (2004), pp 442-452. https://doi.org/10.1002/jnr.20083

- 36. Tian, H. and Ma, M., "Molecular organization of the olfactory septal organ", Journal of Neuroscience, vol.24, (2004), pp 8383-8390. https://doi.org/10.1523/JNEUROSCI.2222-04.2004
- J. Fleischer, K. Schwarzenbacher, S. Besser, N. Hass and H. Breer, "Olfactory receptors and signaling elements in the Grueneberg ganglion", Journal of Neurochemistry, vol 8, (2006), pp 543-54. https://doi.org/10.1111/j.1471-4159.2006.03894.x
- 38. Carlson, J. R., "Olfaction in Drosophila: from odor to behavior", Trends in Genetics, vol 12, (1996), pp 175-180.
- 39. Mori, I., "Genetics of chemotaxis and thermotaxis in the nematode Caenorhabditis elegans", Annual Review of Genetics, vol 33, (1999), pp 399-422. https://doi.org/10.1146/annurev.genet.33.1.399
- 40. Yoshihara, Y., "Zebrafish olfactory system", In: Mori, K.(eds) The olfactory system; Springer, Tokyo, (2014). https://doi.org/10.1007/978-4-431-54376-3_5
- Bruyne, M. D., Clyne, P. J. and Carlson J. R., "Odor coding in a model olfactory organ: the Drosophila maxillary palp", Journal of Neuroscience, vol 19, (1999), pp 4520-4532. DOI: https://doi.org/10.1523/jneurosci.19-11-04520.1999
- 42. De Bruyne, M., Foster, K. and Carlson, J. R., "Odor coding in the Drosophila antenna", Neuron, vol 30, (2001), pp 537-552. https://doi.org/10.1016/s0896-6273(01)00289-6
- 43. Vosshall, L. B., "Olfaction in Drosophila", Current Opinion in Neurobiology, vol 10, (2000), 498–503. https://doi.org/10.1016/s0959-4388(00)00111-2
- Martin, F., Boto, T., Gomez-Diaz, C. and Alcorta, E., "Elements of olfactory reception in adult Drosophila melanogaster", Anatomical Record (Hoboken, N. J.: 2007), vol 296, (2013), pp 1477–1488. https://doi.org/10.1002/ar.22747
- 45. Stocker, R. F., "The organization of the chemosensory system in Drosophila melanogaster: a review", Cell and Tissue Research, vol 275, (1994), pp 3–26. https://doi.org/10.1007/BF00305372
- 46. Spehr, M. and Munger, S. D., "Olfactory receptors: G protein-coupled receptors and beyond", Journal of Neurochemistry, vol 109, (2009), pp 1570–1583. https://doi.org/10.1111/j.1471-4159.2009.06085.x
- Akhila, A., "Metabolic engineering of biosynthetic pathways leading to isoprenoids: Mono- and sesquiterpenes in plastids and cytosol", Journal of Plant Interactions, vol 2, (2007), pp 195–204. https://doi.org/10.1080/17429140701670953
- Maffei, M. E., Gertsch, J. and Appendino, G., "Plant volatiles: production, function and pharmacology", Natural Product Reports, vol 28, (2011), pp 1359–1380. https://doi.org/10.1039/c1np00021g
- 49. Buck L. and Richard Axel, A., "A novel multigene family may encode odorant receptors: a molecular basis for odor recognition", Cell, vol 65, (1991), pp 175-187.
- 50. Godfrey, P. A., Bettina, M. and Buck, L. B., "The mouse olfactory receptor gene family", Proceedings of the National Academy of Sciences, vol 101, (2004), pp 2156-2161. https://doi.org/10.1073/pnas.0308051100
- Sullivan, S. L., Adamson, M. C., Ressler, K. J., Kozak, C. A. and Buck, L. B., "The chromosomal distribution of mouse odorant receptor genes", Proceedings of the National Academy of Sciences, vol 93, (1996), pp 884-888. https://doi.org/10.1073/pnas.93.2.884
- Niimura Y. and Masatoshi, N., "Evolutionary dynamics of olfactory and other chemosensory receptor genes in vertebrates", Journal of Human Genetics, vol 51, (2006), pp 505-517. https://doi.org/10.1007/s10038-006-0391-8
- 53. Zhang, X., Rodriguez, I., Mombaerts, P. and Firestein, S., "Odorant and vomeronasal receptor genes in two mouse genome assemblies", Genomics, vol 83, (2004), pp 802–811.

https://doi.org/10.1016/j.ygeno.2003.10.009

- 54. Rouquier, S., Taviaux, S., Trask, B. J., Brand-Arpon, V., Engh, G. V. D., Demaille, J. and Giorgi, D., "Distribution of olfactory receptor genes in the human genome", Nature Genetics, vol. 18, (1998), pp 243-250. https://doi.org/10.1038/ng0398-243
- 55. Glusman, G., Yanai, I., Rubin, I. and Lancet, D., "The complete human olfactory subgenome", Genome Research, vol 11, (2001), pp 685-702.
- 56. Ngai, J., Dowling, M. M., Buck, L., Axel, R. and Chess, A., "The family of genes encoding odorant receptors in the channel catfish", Cell, vol 72, (1993), pp 657-666.
- 57. Freitag, J., Krieger, J., Strotmann, J. and Breer, H., "Two classes of olfactory receptors in Xenopus laevis", Neuron, vol 15, (1995), pp 1383-1392.
- 58. Niimura, Y. and Nei, M., "Extensive gains and losses of olfactory receptor genes in mammalian evolution", PLOS One, vol 2, (2007), pp e708. https://doi.org/10.1371/journal.pone.0000708
- 59. Malnic, B., Gonzalez-Kristeller, D. C. and Gutiyama, L. M., "Odorant Receptors", In: The neurobiology of olfaction. Boca Raton (FL): CRC Press/Taylor & Francis, vol 7, (2010), pp 181-202. https://www.ncbi.nlm.nih.gov/books/NBK55985/
- 60. Maresh, A., Rodriguez, G. D., Whitman, M. C. and Greer, C. A., "Principles of glomerular organization in the human olfactory bulb – implications for odor processing", PLOS One, vol 3, (2008), pp e2640. https://doi.org/10.1371/journal.pone.0002640
- 61. Gilad, Y., Rifkin, S. A., Bertone, P., Gerstein, M. and White, K. P., "Multi-species microarrays reveal the effect of sequence divergence on gene expression profiles", Genome Research, vol 15, (2005), pp 674-680.
- 62. Tirindelli, R., Dibattista, M., Pifferi, S. and Menini, A., "From pheromones to behavior", Physiological Reviews, vol 89, (2009), pp 921-956. https://doi.org/10.1152/physrev.00037.2008
- Fu, X., Zhou, Y., Zeng, L., Dong, F., Mei, X., Liao, Y., Watanabe. N. and Yang. Z., "Analytical method for metabolites involved in biosynthesis of plant volatile compounds", RSC Advances, vol 7, (2017), pp 19363-19372. https://doi.org/10.1039/C7RA00766C
- 64. Pichersky, E., Noel, J. P. and Dudareva, N., "Biosynthesis of plant volatiles: nature's diversity and ingenuity", Science, vol 311, (2006), pp 808-811. https://doi.org/10.1126/science.1118510
- 65. Hsieh, M. H., Chang, C. Y., Hsu, S. J. and Chen, J. J., "Chloroplast localization of methylerythritol 4phosphate pathway enzymes and regulation of mitochondrial genes in ispD and ispE albino mutants in Arabidopsis", Plant Molecular Biology, vol 66, (2008), pp 663-673.https://doi.org/10.1007/s11103-008-9297-5
- 66. Simkin, A. J., Guirimand, G., Papon, N., Courdavault, V., Thabet, I., Ginis, O., Bouzid, S., Giglioli-Guivarc'h N. and Clastre, M., "Peroxisomal localization of the final steps of the mevalonic acid pathway in planta", Planta, vol 234, (2011), 903-914. https://doi.org/10.1007/s00425-011-1444-6
- 67. Pulido, P., Perello, C. and Rodriguez-Concepcion, M., "New insights into plant isoprenoid metabolism", Molecular Plant, vol 5, (2012), pp 964-967.
- 68. McGarvey, D. J. and Croteau, R., "Terpenoid metabolism", The Plant Cell, vol 7, (1995), pp 1015. https://doi.org/10.1105%2Ftpc.7.7.1015
- 69. Cane, D. E., "Sesquiterpene biosynthesis: cyclization mechanisms", Comprehensive Natural Products Chemistry, vol 2, (1999), pp 155-200.
- 70. Knudsen, J. T., Eriksson, R., Gershenzon, J. and Ståhl, B., "Diversity and distribution of floral scent", The Botanical Review, vol 72, (2006), pp 1-20. https://doi.org/10.1663/0006-8101(2006)72 [1: DADOFS]2.0.CO;2

- 71. Maeda, H. and Dudareva, N., "The shikimate pathway and aromatic amino acid biosynthesis in plants", Annual review of plant biology, vol 63, (2012), pp 73-105. https://doi.org/10.1146/annurev-arplant-042811-105439
- 72. Feussner I. and Wasternack, C., "The lipoxygenase pathway", Annual Review of Plant Biology, vol 53, (2002), pp 275-297. https://doi.org/10.1146/annurev.arplant.53.100301.135248
- 73. Miller P., "The Gardeners Dictionary: Containing the Best and Newest Methods of Cultivating and Improving the Kitchen, Fruit, Flower Garden, and Nursery A-L. Author, (1759).
- 74. Palen, G. F. and Goddard, G. V., "Catnip and oestrous behaviour in the cat", Animal Behaviour, vol 14, (1966), pp 372-377. https://doi.org/10.1016/S0003-3472(66)80100-8
- 75. Todd, N. B., "The catnip response", Ph. D. Thesis. Harvard University, Cambridge, Massachusetts, (1963).
- 76. Hill, J. O., Pavlik, E. J., Smith, G. L., Burghardt G. M. and Coulson, P. B., "Species-characteristic responses to catnip by undomesticated felids", Journal of Chemical Ecology, vol 2, (1976), pp 239-253. https://doi.org/10.1007/BF00987747
- 77. Bates, R. B., Eisenbraun, E. J. and McElvain, S. M., "The configurations of the nepetalactones and related compounds", Journal of the American Chemical Society, vol 80, (1958), pp 3420-3424. https://doi.org/10.1021/ja01546a054
- Yoshii, N., Hano, K. and Suzuki, Y., "Effect of certain substances isolated from 'matatabi' on the EEG of cat", Psychiatry and Clinical Neurosciences, vol 17, (1963), pp 335-350. https://doi.org/10.1111/j.1440-1819.1963.tb00003.x
- 79. Jackson, B. and Reed, A., "Catnip and the alteration of consciousness", Journal of the American Medical Association, vol 207, (1969), pp 1349-1350. doi:10.1001/jama.1969.03150200115019
- Hart, B. L. and Leedy, M. G., "Analysis of the catnip reaction: mediation by olfactory system, not vomeronasal organ", Behavioral and Neural Biology, vol 44, (1985), pp 38-46. https://doi.org/10.1016/S0163-1047(85)91151-3
- Campbell, J. F., Bindra, D., Krebsand, H. and Ferenchak, R. P., "Responses of single units of the hypothalamic ventromedial nucleus to environmental stimuli", Physiology & Behavior, vol 4, (1969), pp 183-187. https://doi.org/10.1016/0031-9384(69)90078-X
- 82. Todd, N. B., "Inheritance of the catnip response in domestic cats", Journal of Heredity, vol 53, (1962), pp 54-56.
- Katahira, K. and Iwai, E., "Effect of unilateral lesion of amygdala on unmanifested response to matatabi (Actinidia polygama) in cats", The Tohoku Journal of Experimental Medicine, vol 115, (1975), pp 137-143.
- Macdonald, D., "The carnivores: order Carnivora", In: Social odours in mammals, (Eds: R. E. Brown & D.W. Macdonald), Clarendon Press, vol 2, (1985), pp 619–722.
- Scaffidi, A., Algar, D., Bohman, B., Ghisalberti E. L. and Flematti, G., "Identification of the cat attractants isodihydronepetalactone and isoiridomyrmecin from Acalypha indica", Australian Journal of Chemistry, vol 69, (2016), pp 169-73. https://doi.org/10.1071/CH15476
- 86. Indian Patent No. 426217, "A patent for isolating the chemical compounds from the fresh roots of Acalypha indica Linn", (2017), Chemical Signal and Lipidomics Lab, Department of Botany (Centre of Advanced Study), University of Calcutta.
- Mendl, M., Randle K. and Pope, S., "Young female pigs can discriminate individual differences in odours from conspecific urine", Animal Behaviour, vol 64, (2002), pp 97-101. https://doi.org/10.1006/anbe.2002.3040
- 88. Rekwot, P. I., Ogwu, D., Yedipe, E.O. and Sekoni, V.O., "The role of pheromones and biostimulation

in animal reproduction", Animal Reproduction Science, vol 65, (2001), pp 157-170. https://doi.org/10.1016/S0378-4320(00)00223-2

- Rørvang, M.V., Schild, S.L. Wallenbeck, A., Stenfelt, J., Grutand, R., Valros, A. and Nielsen, B. L., "Rub n roll– pigs, sus Scrofa domesticus, display rubbing and rolling behaviour when exposed to odours", Applied Animal Behaviour Science, vol 266, (2023), pp 106022. https://doi.org/10.1016/j.applanim.2023.106022
- 90. Tucker A. O. and Tucker, S. S., "Catnip and the catnip response", Economic Botany, vol 42, (1988), pp 214–231. https://doi.org/10.1007/BF02858923
- 91. Purohit R.M. and Nigam, S. S., "Chemical examination of the essential oil derived from the leaves of Nepeta hindostana Roth", Saugar, India (City) Univ. J. Pt. II, Sect. A. vol 8, (1959), pp 46-49.
- 92. Murai, F., "Chemical components in matatabi. II. The structure of matatabilactone", Journal of Chemical Society, Japan, Pure Chemistry Sect 81 (1960), pp 1324-1326.
- 93. Torssell, K. and Wahlberg, K., "The structure of the principal alkaloid from Valeriana officinalis (L.)", Tetrahedron Letters, vol 7, (1966), pp 445-448. https://doi.org/10.1016/S0040-4039(00)72961-3
- 94. Pagnoni, U. M., Pinetti, A., Trave, R. and Garanti, L., "Monoterpenes of Teucrium marum", Australian Journal of Chemistry, vol 29, (1976), pp 1375-1381. https://doi.org/10.1071/CH9761375
- 95. Sakan, T., Murai, F., Isoe, S., Hyeon S. B. and Hayashi, Y., "The biologically active C9-, C10-, and Cii- terpenes from Actinidia polygama Miq., Boschniakia rossica Hult, and Menyanthes trifoliata L", Nippon Kagaku Zasshi, vol 90, (1969), pp A29-33.
- 96. Jackson, B. and Reed, A., "Catnip and the alteration of consciousness", Journal of American Medical Association, vol 207, (1969), pp 1349–1350. doi:10.1001/jama.1969.03150200115019
- 97. Touhara K. and Vosshall, L.B., "Sensing odorants and pheromones with chemosensory receptors", Annual review of Physiology, vol 71, (2009), pp 307-32. https://doi.org/10.1146/annurev.physiol.010908.163209
- 98. Schild, S. L. and Rørvang, M. V., "Pig olfaction: the potential impact and use of odors in commercial pig husbandry", Frontiers in Animal Science, vol 4, (2023), p 1215206.
- 99. Waller, G. R., Price, G. H. and. Mitchell, E. D., "Feline attractant, cis, trans-nepetalactone: metabolism in the domestic cat", Science (New York), vol 164, (1969), pp 1281–1282. https://doi.org/10.1126/science.164.3885.1281
- 100. Hayashi, T., "Motor reflexes of cats to Actinidiapolygama (Japan) to catnip (USA). Theories of odor and odor measurement", N. N. Tanyolac, Istanbul. (1968), pp 351-358.
- 101. Wickramaratne, S., de Silva, N., Wickramasinghe, S., Wanigasekara, A., Adhikari R. and Rajapakasha, E., "Olfactory behaviour reactions to Acalypha indica preparations in domestic cats", Applied Animal Behaviour Science, vol 257, (2022), p 105776. https://doi.org/10.1016/j.applanim.2022.105776
